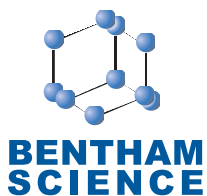


REVIEW ARTICLE

Beta-glucans is a Potential Inhibitor of Ovarian Cancer: Based on Molecular and Biological Aspects



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Abstract: Ovarian cancer is a lethal type of cancer which is initiated to the ovaries and affects 1 out of every 75 women. Due to the high number of deaths (almost 152,000) related to this cancer, it seems that novel efficient therapeutic methods are required in this field. Beta-glucans are a type of glucose linear polymers which have been proven to have a lot of advantageous activities. Recently, investigations have declared that these polysaccharides have the potential to be used as anti-cancer drugs. These agents are able to affect several mechanisms such as inflammation and apoptosis, and that is how cancers are prone to be affected by them. In this review, we attempt to investigate the role of beta-glucans on ovarian cancer. We hope that this paper would give some novel insights into the field of ovarian cancer treatment.

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1. INTRODUCTION

Ovarian cancer (OC) is one of the most lethal gynecological cancers, which mostly occurs in women at the age of 40 or more [1]. This cancer takes 152,000 lives every year and thus, it is acknowledged as the eighth frequent cancer which is causing death among women of the world [2, 3]. It is estimated that the risk of being affected by ovarian cancer is 1 in 75 for every woman and 1 woman patient out of 100 is being killed by this cancer [2]. The survival rate of this cancer does not exceed from 45% and the number of new women diagnosed is augmenting every year [1, 2]. According to evidence, a set of various cancers with diverse origins (cellular and anatomical), pathogenesis, alterations in molecular mechanisms, and expression of oncogenic genes are acknowledged as OC. Between all of these cancers, carcinomas and between the epithelium-related cancers (including serous, endometrioid, clear cell and mucinous), high-grade serous carcinoma are the most common ovarian cancer subtypes [2, 4, 5]; thus, in this paper, our main focus will be on ovarian carcinomas.

Being impacted by this cancer can be more likely in women who have high body mass index (BMI), the habit of

smoking and alcohol drinking, and infertility. Furthermore, some other risk factors such as pregnancy, lactation, oophorectomy, consumption of oral contraceptives, and exercise are approved to have a reverse effect on the ovarian cancer risk [2, 6, 7]. After detecting some symptoms in a patient, some procedures are confirmed to be effective for a certain diagnosis. These procedures encompass testing the amounts of serum glycoprotein CA125, computed tomography or CT scan, magnetic resonance imaging or MRI, and ultrasonography [8].

From the treatment perspective, the first step is staging and determining the spread of cancer [8]. In general, four stages account for this cancer which should be managed through different therapeutic strategies [9]. The most common method utilized for the treatment of early stages is surgery [8]. Noteworthy, surgical management relies on the exact stage of the disease and contains hysterectomy and uni or bilateral salpingo-oophorectomy [8]. Nevertheless, using surgery alone is rarely reported to be effective in advanced stages of OC [10]. Intravenous and intraperitoneal chemotherapy are two convenient methods (adjuvant to surgery) in confronting a patient with a stage 2 and above ovarian malignancy [10, 11]. Still and all, platinum-based cytotoxic drugs such as paclitaxel are not being used for patients with recurrent ovarian cancer [8]. Neoadjuvant chemotherapy or NACT is another therapeutic approach which its usage is increased recently [9, 12, 13]. In this method, chemotherapy should be administered

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before operating any surgery [9, 12]. Besides, there are also some other options for OC treatment, including radiation therapy, PARP inhibiting, immunotherapy including PD-1/PD-L1-targeted immune checkpoint inhibitors and VEGFR inhibitors, targeting some antigens and signaling pathways related to OC including PI3K (Phosphatidylinositol 3-Kinase), folate receptor alpha, and Protein tyrosine kinase 7 which are mostly being used in advanced and metastatic stages and the recurrence of this disease [14-19]. All taken together, searching for novel drugs, agents, or methods are needed for enhancing the status of OC patients and reducing the mortality and morbidity rates of this fatal cancer. In such wise, we tend to investigate the advantageous effects of the family of beta-glucans on the cancerous cells especially the ones are located in the ovaries, fallopian tube, uterine ligaments (and adnexa), and the peritoneum and retroperitoneum spaces which all are being known in the name of ovarian cancer [20].

2. WHAT ARE BETA-GLUCANS?

Glucose molecules are able to bind to each other with diverse kinds of linkages and create various dimers, oligomers, and polymers. When a great number of D-glucose monomers are linked together with several beta-glycosidic bonds, a polymer is established that is called "beta-glucan" [21]. These natural polysaccharides are mainly composed of a linear core chain with β (1 \rightarrow 3) bonds and a number of branches which are linked to the core chain by β (1 \rightarrow 4) or β (1 \rightarrow 6) linkages [22]. Mostly, there are three sources for extraction of beta-glucans: plants such as oat and barley [23, 24], fungi such as yeasts and Shiitake mushrooms [25, 26], and bacteria such as *Agrobacterium* [27]. These polymers take numerous parts in our body by dint of attaching to their cellular receptors. In our knowledge, four types of receptors are able to provide a connection with these molecules.

Complement receptor 3 or CR3 is one of the most important receptors that allows beta-glucans to function [28]. This receptor is composed of two chains: α and β [29]. The alpha chain is a kind of CD11 that is known to have a, b, and c types, and beta chain is a CD18 protein [22, 28]. This receptor is mostly found in the cellular membrane of myeloid-derived cells such as natural killer cells, macrophages, neutrophils, and dendritic cells as a transmembrane glycoprotein [30]. CR3 has the ability to bind to a beta-glucan by its α_M chain and bind to one of the ingredients of complement (iC3b) from another site [29]. CR3 has the possibility of binding to a great number of endogenous and exogenous ligands and, after creating the connection, it facilitates the complement-associated opsonization and finally triggers the phagocytosis by immunity cells [31, 32]. Dectin-1 is another essential receptor for beta-glucans to act upon [33, 34]. This receptor is a member of C-type lectin family, which comprises extracellular (C-terminal), transmembrane (a single domain), and intracellular (amino-terminal) components [35, 36]. Dectin-1 expression is in similar cells with CR3 receptor which have a myeloid ancestor [37]. Moreover, this receptor is secondarily expressed in T and B lymphocytes [37]. Many functions are attributed to this class of receptors, such as inducing phagocytosis, increasing

the release of inflammatory mediators such as cytokines and chemokines, and increasing the oxidative burst conducted by neutrophils [28, 36, 38, 39].

Besides the mentioned receptors, there are also some other additional receptors whose expressions and functions are lesser in amount and number in comparison to CR3 and dectin-1 [36]. Lactosylceramide is one of these receptors that can be found on the surface of the polymorphonuclear leukocytes in humans [40]. Similar to the mentioned tasks of CR3 and dectin-1, this receptor is also able to augment cytokine secretion and oxidative burst after binding to beta-glucans [40, 41]. CD5, which is a scavenger receptor, is another member of the beta-glucan receptor group [42]. The cellular membranes of mature T and B1a cells are where CD5 can be found [42]. The exact tasks and mechanisms of this receptor are not clear yet, but generally, it has the ability to inhibit the activity of B cells signaling, increase the secretion of interleukin-8, and activate ERK and p38 MAP kinase in Jurkat T cells [29, 42, 43].

3. OVARIAN CANCER PATHOGENESIS

According to investigations on the cause of OC, it seems that more than 23% of ovarian cancers are inherited [44]. There are several genetic abnormalities which are known to be responsible for hereditary OC, including BRCA1/2, TP53, BARD1, CHEK2, and RAD51 which BRCA1 and 2 seem to be the most common among them [44]. Carriers of BRCA1 and 2 mutations have a higher chance of getting OC (BRCA1 40%–60% and BRCA2 11%–30%) and in about 65–85% of OC patients, these mutations are detectable [45, 46].

After all, the main question is "how does BRCA mutation result in ovarian cancer?". The answer to this question relies on the DNA damage responses, which get help from these proteins to detect and repair the DNA lesions [47]. Both of these proteins are involved in repairing the double-stranded lesions of DNA, which is mostly possible through non-homologous end-joining (NHEJ) and homologous recombination (HR) repair pathways [47]. In the HR process, BRCA1 is recruited to the damage site by histone H2AX and takes part in DNA end resection by means of CtIP ubiquitination, which initiates the nuclease activities on DNA strands [48, 49].

BRCA2 is also a mediator of HR, aiding the process of RPA removal from the single-stranded DNA, which is established in the DNA end resection stage of HR [48]. Therefore, BRCA2 is participating in the generation of a RAD51-related nucleoprotein filament [49].

Overall, BRCA1/2 mutations or epigenetic inactivation cause the deficiency of HR pathway in ovarian cells and increase their susceptibility to genome defects which is one of the bases of tumor initiation [50].

However, despite all of our improvements in the field of cancer pathogenesis, still, an exact answer is missing for the questions about the pathogenesis of ovarian cancer. Investigators have represented some hypotheses by means of cellular and molecular pathways, which we discuss in this section separately.

3.1. Inflammation

During the process of inflammation, immune cells are recruited at the site of injury or infection due to the secretion of some cytokines and chemokines [51]. Destruction of the injured cells or pathogenic agents is being conducted by some cells such as macrophages, dendritic cells, and neutrophils and some mechanisms such phagocytosis and the release of free radicals, TNF- α , interleukins, serine and cysteine proteases, and interferons [52]. After the binding of inflammatory mediators to a cell, some signaling pathways are triggered, including NF- κ B or nuclear factor kappa-light-chain-enhancer of activated B cells, MAPKs or p38 mitogen-activated protein kinases, and JNK or c-Jun N-terminal kinase [53-55]. Recently, a great body of evidence has revealed the role of different components of this process in the initiation of many diseases such as cancer [56]. Mutating cellular DNA and triggering NF- κ B pathway are two main known reasons why inflammation leads to cancer [57]. In ovarian cancer perspective, inflammation is reported to be one of the main pathogenesis-related mechanisms by many researchers [58]. One of the most reliable hypotheses in this field is “incessant ovulation” [58]. It is believed that every month during ovulation, ovarian epithelium undergoes a bunch of injuries which is being repeated in the entire life of a woman, and this occurrence results in induced inflammatory responses [57, 58].

The role of inflammation in each step of ovulation is confirmed; for instance, Luteinizing hormones are causing an augmentation in the number of neutrophils in the ovaries before the ovulation [57]. Moreover, there are several macrophages in the thecal layer of ovaries which are responsible for secreting epidermal growth factor (EGF), reactive oxygen species or ROS, tumor necrosis factor-alpha (TNF- α), and interleukin-1 β (IL-1 β). Increased amounts of these mediators manage some alterations in the expression of some inflammatory genes and induce cyclooxygenase-2 (COX-2) and IL-8 expression [57, 59, 60]. In addition to the ovulation process itself, there are also some other factors that are able to boost the risk of OC through inflammation. These factors include infection in the reproductive system, overweight, endometriosis, and polycystic ovarian syndrome (PCOS) [61-65]. Overall, evidence demonstrates that inflammation is able to help ovarian cancer initiation by two mechanisms: secreting cytokines, chemokines, and growth factors and releasing free radicals such as ROS and providing oxidative stress.

3.2. Apoptosis

The lack of apoptosis or programmed cell death is one of the fundamental parts of cancer pathogenesis [66]. There are two main pathways that provoke the process of apoptosis: intrinsic and extrinsic [67]. The former pathway is conducted by mitochondria and their released agents and the latter pathway is conducted through the binding of some ligands such as FasL to their receptors on the cell surface [67]. Both of these pathways have the task of activating some cysteine proteases named caspases [68]. These proteases are responsible for most of the alterations happening in an apoptotic cell [68]. Additionally, there are some genes and their related proteins like Bcl-2 and p53, which are regulating

apoptosis by triggering or inhibiting it in times of need [67]. Overall, any disturbance in any ingredient of this process is able to establish a disease like malignancies.

Free radicals like oxygen or nitrogen reactive species are one of the factors that have the ability to activate some signaling pathways, such as HIF-1 α and thus, disturb the balance between apoptosis and proliferation in a cell. Some cellular mechanisms exist which have the duty of inhibiting free radicals; for instance, N(G)-nitro-L-arginine methyl ester or L-NAME is an inhibitor of nitric oxide (NO) synthesis and prevents the accumulation of NOS in cells [69].

In the case of OC, investigations express that apoptosis helps the cancer progression in two ways: promoting apoptosis in lymphocytes, which is the cause of IL-6 secretion, and the suppression of immune system [70]. Although the main mechanism happens in the cancerous cells: in these cells, L-NAME (N(G)-nitro-L-arginine methyl ester) inhibits iNOS and therefore, apoptosis is down-regulated in these cells because of NO/iNOS pathways [71].

3.3. Uncontrolled Cell Proliferation

Uncontrolled cell proliferation is one of the most essential cancer hallmarks, which is mostly triggered in cancerous ovarian cells by inflammation-associated signaling pathways. PI3K/AKT/mTOR is one of these pathways which aids tumor formation through a diversity of mechanisms, including increasing cellular proliferation [72]. Targeting ingredients of this signaling pathway by specific siRNAs has shown that the proliferation of cancerous cells of ovaries can be suppressed [73, 74]. On the other hand, NF- κ B is also an essential part of OC pathogenesis which affects the cell cycle by regulating its checkpoints, activating mitogen-activated protein kinase (MAPK) phosphorylation, and regulating mortalin [75-77]. On the other hand, it might also be related to uncontrolled proliferation of OC cells. Guan *et al.*, [78] found out that higher levels of PUM1 gene (pumilio RNA binding family member 1 in these cells is involved in cancer initiation). This gene is associated with chromosomal mutations [78].

Additionally, there are some other genes whose abnormal expression is observed to be related to increased proliferation of OC cells and their effect is approved after observing the suppressed proliferation when they are targeted by non-coding RNAs. PTEN, PLXNB2, YWHAZ, hTERT, FOXA1, SNHG17, S100A10, PTCH1, and ZEB1 are some of these genes [79-86].

4. DIVERSE FUNCTIONS OF BETA-GLUCANS IN CANCER

In general, a wide range of functions are attributed to beta-glucans but what is orienting many researchers towards them is their anti-cancer and immunologic effects. Inflammation, oxidative stress, and apoptosis are three main mechanisms that are being affected by these polysaccharides (summarized in Fig. 1), but before discussing these mechanisms, we would take a brief look at beta-glucan's cellular effects.

The cellular impact of these polymers is mediated by different types of membrane receptors containing Dectin-1, complement receptor 3 or CR3, lactosylceramide, and Toll-like receptors [87]. Dectin-1 receptor is mostly expressed at

the surface of inflammatory cells, including neutrophils, macrophages, and dendritic cells. Dectin-1/Syk/NF- κ B signaling is one of the most important axes through which this receptor works [88]. NF- κ B activation is the share point of Dectin-1 and lactosylceramide receptor which participates in pro-inflammatory responses [89]. Similar to Dectin-1, CR3 is also broadly expressed on macrophages and natural killer cells and, when activated by beta-glucans, binds to a variety of cells encompassing antibody-coated cancer cells [90].

4.1. Inflammation

With respect to researches, beta-glucans are affecting both innate and adaptive immunities and thereby modulate the inflammatory response [91]. The influence of beta-glucans on inflammation is possible in two ways: affecting the production of cytokines/chemokines and impacting the proliferation and activation of immune cells. Some studies have approved the former mechanism; for example, Olson *et al.*, showed that beta-glucans are increasing the release of TNF- α by neutrophils [92]. Increasing the production of TNF- α by macrophages is also detected due to the binding of beta-glucans to CR3, Dectin-1, and TLR-2 [91]. In addition to TNF- α , serum levels of some other cytokines such as IL-1 α , IL-2, IL-4, IL-8, IL-10, and IFN-gamma are investigated after beta-glucan administration [93-95].

The latter mechanism or affecting immune cells such as dendritic cells, Myeloid-derived suppressor cells, macrophages, neutrophils, and leukocytes are also explored by a great body of research [40, 95-98]. In macrophages, regulating Phospholipase A2 and increasing the cyclooxygenase 2 expression is achievable after the administration of zymosan [99]. Zymosan increases arachidonic acid and leukotriene C4 through Dectin-1 and increases COX2 expression and prostaglandin through TLR2 [99]. Zymosan also affects leukocytes in a direct manner and, after activating them, stimulates phagocytosis and the amounts of ROS secreting from them [100]. In view of leukocyte, "beta-glucans shift the tumor microenvironment toward a T cell-sensitive environment" [101].

4.2. Oxidative Stress

Wakshull *et al.* revealed that beta-glucans are able to improve the oxidative burst response of neutrophils by binding to one of their receptors: lactosylceramide [40]. Furthermore, the interaction between beta-glucans and C-type lectin receptors (CLRs) is widely reviewed by Goyal *et al.* [102]. CLRs are known for their ability to bind to carbohydrates and include several receptors like Dectin-1 and Dectin-2. It seems that in addition to immune cells, pulmonary epithelial cells are also prone to be affected by beta-glucans via Dectin-1 and afterward, generate ROS [103]. Liang *et al.* [104] also observed an augmentation in oxidative burst response conducted by neutrophils in rats after PGG-Glucan treatment. They also revealed that this effect is not dependent on the expression of TNF- α and IL-1 β in rats [104]. Bose *et al.* tried to reveal the exact receptor which mediates this function of beta-glucans in monocytes [105]. They used GE2 antibody for blocking Dectin-1 and a combination of antibodies for different subunits of CR3. They concluded that Dectin-1 inhibition effectively decreases the SO production in

these cells while CR3 inhibition doesn't affect oxidative burst [105].

4.3. Apoptosis

There are not many papers directly mentioning apoptosis as an important influence of beta-glucans, but beta-glucans can indirectly affect apoptosis in cancer cells. For instance, affecting NF- κ B pathway by these agents is related to apoptosis alterations [106, 107]. Another way of inducing apoptosis is by increasing oxidative stress [108], which we have declared before. Sensitizing cancerous cells to apoptosis by means of caspase 8 and 3 activation is one of the mechanisms by which these agents work [109, 110]. In addition, transferring BID into the mitochondria is also stimulated by beta-glucans which results in apoptosis [109]. Beta-glucans are also capable of regulating apoptosis-related genes, such as decreasing the expression of Bcl-2 and increasing the expression of Bax [110, 111]. On the other hand, β -D-glucans also induce necrosis along with apoptosis which inhibits the progression of cancer cells [112].

5. HAVE BETA-GLUCANS EVER BEEN USED IN CANCER TREATMENT?

A great body of evidence has shown how beta-glucans have the potential to help overcoming cancer. For instance, Thomas *et al.* used PGG in combination with chemotherapy for the treatment of advanced lung cancer [113]. Furthermore, there is also some other evidence for lung cancer [114]. Ostadrahimi *et al.* [115] revealed that using beta-glucans adjuvant to chemotherapy is useful for breast cancer treatment. Kataoka *et al.* revealed the roles of lentinan on the life quality of gastric cancer patients [116]. Tari and colleagues also worked on lentinan and found that it is useful for the treatment of metastatic prostate carcinoma [117]. Overall, these evidence (and a great number of others that we have not mentioned) demonstrate that beta-glucans have the potential for being administered adjuvant to chemotherapy in order to enhance their efficacy and decrease their side effects on the patients of different cancers.

6. BETA-GLUCANS AND OVARIAN CANCER: MECHANISMS

In regard to the many roles that beta-glucans have in cancer treatment, they might have some advantages in ovarian cancer treatment, as well. Therefore, we gathered some evidence to investigate the exact functions of these natural polymers in OC (Fig. 1).

6.1. Lentinan

Lentinan is a β -(1 \rightarrow 3)-d-glucan that can be excreted from a common edible mushroom, *Lentinus edodes* [118]. A case-control trial on an OC patient with recurrent cancer in the lymph nodes declared that Adoptive immunotherapy with lentinan without using any chemotherapeutic drugs is useful for the treatment of lymph node metastases from ovarian cancer [119]. Another case-control study also revealed that utilizing intravenous lentinan with IA 5FU increases the production of IL-6 and, overall, inhibits cancer recurrence [120]. Another paper also confirmed the anti-cancer effects of

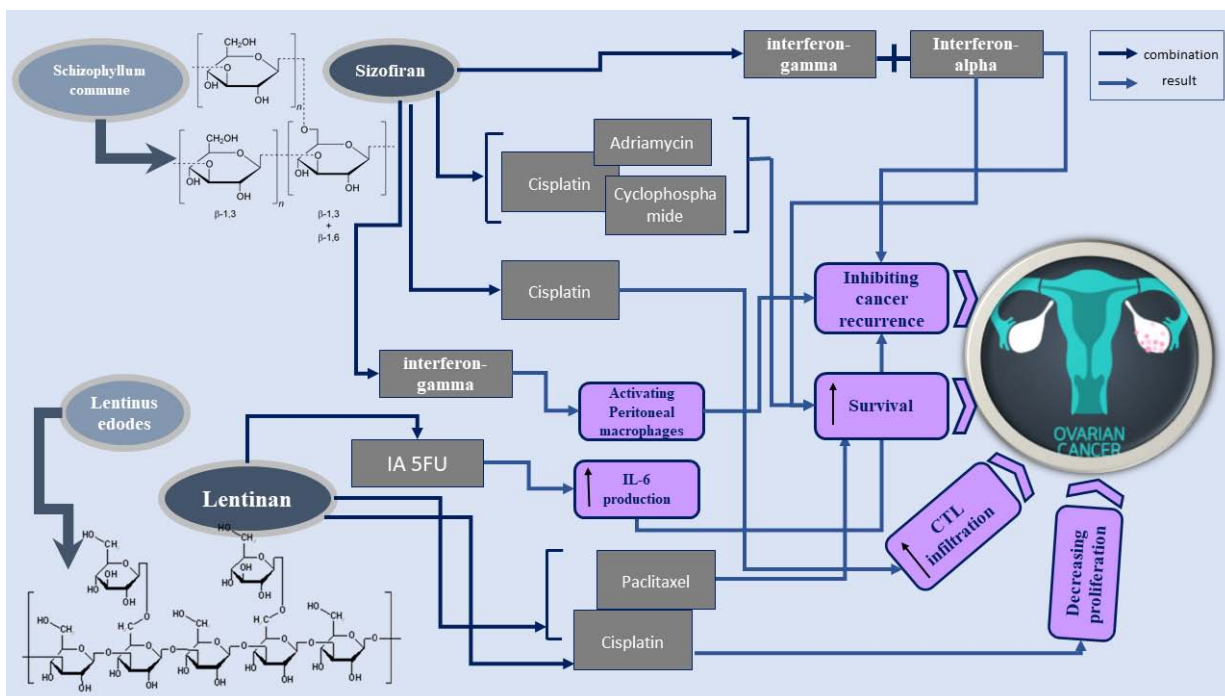


Fig. (1). Schematic representation of how beta-glucans exerts its functions in ovarian cancer. (A higher resolution/colour version of this figure is available in the electronic copy of the article).

lentinan on advanced ovarian cancer patients by combining this beta-glucan with cisplatin and paclitaxel [121, 122].

Furthermore, using the combination of lentinan and cisplatin on Hey ovarian cancer cells inhibited these cells by decreasing their proliferation [123] (Fig. 1).

6.2. Sizofiran

Schizophyllan or SPG is produced by the fungus *Schizophyllum commune*. Schizophyllan is a β -1, 3 beta-glucan with β -1, 6 branching. Schizophyllan is also known as sizofiran [124]. Chen *et al.* [125] used combined immunotherapy on 12 ovarian cancer patients and found that the combination of sizofiran with Interferon-alpha and interferon-gamma is helping the survival of OC patients and no recurrence was observed in these patients. Hoshino and colleagues used another combination (sizofiran and rG-CSF) on OC patients who are going through chemotherapy. They revealed that this combination could be useful for myelosuppression by chemotherapy for ovarian cancer [126]. Another group of investigators also worked on this beta-glucan and revealed that utilizing sizofiran or SPG in patients undergoing chemotherapy with cisplatin, adriamycin and cyclophosphamide can enhance the prognosis in these patients [127]. Sugiyama *et al.* used this agent with cisplatin on OC rats and demonstrated that this combination improves the survival rate and cytotoxic T-lymphocyte (CTL) infiltration [128]. Chen *et al.* conducted an *in vivo* trial on 19 patients and expressed that sizofiran and recombinant interferon- γ prevent the recurrence of gynecological cancers especially ovarian cancers, by activating peritoneal macrophages [129] (Fig. 1).

6.3. Cellulose

Cellulose is another beta-glucan that has been used for designing nanoparticles in the field of drug delivery. There are

some studies confirming the efficacy of cellulose-nanoparticles in OC patients, but there is no evidence of direct impacts of this beta-glucan on ovarian cancer cells [130, 131]. However, there is one study using cellulose as a delivery system for cisplatin. Using conjugates of cellulose, 2, 3-dicarboxycellulose (DCC) increases the time of drug release in the cancer site and thus, increases the efficacy of this drug [132].

6.4. Laminarin

Laminarin is beta-1,3-glucan, which can be extracted from brown algae [133]. A recent study has shown some benefits of this beta-glucan against OC cells. *In vitro* investigation of laminarin indicates that it decreases the proliferation of OC cells dose-dependently [134]. Increasing ROS generation, cleaved caspase-3, and caspase 9 are the pro-apoptotic effects of this agent. Furthermore, it is able to trigger mitochondrial apoptosis by increasing the concentration of Ca [134]. More importantly, "laminarin significantly decreased the phosphorylation of PI3K/MAPK signaling proteins in OC cells." All these results suggest that further investigations on this beta-glucan might be useful for providing more therapeutic options for advanced OC [134].

CONCLUSION

Ovarian cancer is one of the most lethal and important health problems among women in the world. The most common treatment used for this cancer is chemotherapy which has a great number of disadvantages on patients. For instance, Chemotherapy-induced peripheral neuropathy (CIPN), Chemotherapy-induced nausea and vomiting (CINV), cognitive dysfunction, mood disorders, fatigue, and cytopenias are the most frequent side-effects between OC patients undergoing chemotherapy (as reviewed by Gockely

[135]). Thus, despite all the improvements in diagnosis and treatment of this cancer, still, more methods and strategies are required for reducing the burden of this disease, increasing its survival rate, and enhancing the life quality of survived patients. Beta-glucans are a kind of glucose polymers that have attracted a lot of attention for their anti-cancer effects and non-toxicity. The role of these linear polysaccharides is investigated in several types of cancers. In this paper, we gathered some evidence on the efficacy of these agents on ovarian cancer. We found that lentinan and sizofiran are useful for enhancing the treatment, preventing recurrence, and decreasing the cytotoxic effects of chemotherapeutic drugs. Although, still more investigations are needed. We suggest that other beta-glucans such as curdlan might also be useful for ovarian cancer treatment (Fig. 1). Moreover, recently some explorations have shown the effectiveness of oral administration of beta-glucans with monoclonal antibodies in the field of cancer treatment [136]. However, there is no evidence for the effectiveness of this method on ovarian cancer. Therefore, we think that working on this subject might give us new insights for OC treatment.

AUTHORS' CONTRIBUTIONS

FS, ZA and JH contributed to the conception, design and drafting of the manuscript. MAM, BY contributed to reviewing relevant literature. All authors approved the final version for submission.

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CONFLICT OF INTEREST

The authors declare no conflict of interest, financial or otherwise.

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REFERENCES

- [1] Webb, P.M.; Jordan, S.J. Epidemiology of epithelial ovarian cancer. *Best Pract. Res. Clin. Obstet. Gynaecol.*, **2017**, *41*, 3-14. <http://dx.doi.org/10.1016/j.bpobgyn.2016.08.006> PMID: 27743768
- [2] Reid, B.M.; Permuth, J.B.; Sellers, T.A. Epidemiology of ovarian cancer: A review. *Cancer Biol. Med.*, **2017**, *14*(1), 9-32. <http://dx.doi.org/10.20892/j.issn.2095-3941.2016.0084> PMID: 28443200
- [3] Kossai, M.; Leary, A.; Scoazec, J-Y.; Genestie, C. Ovarian cancer: A heterogeneous disease. *Pathobiology*, **2018**, *85*(1-2), 41-49. <http://dx.doi.org/10.1159/000479006> PMID: 29020678
- [4] McCluggage, W.G. Morphological subtypes of ovarian carcinoma: A review with emphasis on new developments and pathogenesis. *Pathology*, **2011**, *43*(5), 420-432. <http://dx.doi.org/10.1097/PAT.0b013e328348a6e7> PMID: 21716157

- [5] Prat, J. Ovarian carcinomas: Five distinct diseases with different origins, genetic alterations, and clinicopathological features. *Virchows Arch.*, **2012**, *460*(3), 237-249. <http://dx.doi.org/10.1007/s00428-012-1203-5> PMID: 22322322
- [6] Permuth-Wey, J.; Sellers, T.A. Epidemiology of ovarian cancer. *Methods Mol. Biol.*, **2009**, *472*, 413-437. http://dx.doi.org/10.1007/978-1-60327-492-0_20 PMID: 19107446
- [7] Weiderpass, E.; Botteri, E. Ovarian cancer mortality trends: Which factors are involved? *Ann. Oncol.*, **2016**, *27*(11), 1977-1978. <http://dx.doi.org/10.1093/annonc/mdw411> PMID: 27793848
- [8] Rooth, C. Ovarian cancer: Risk factors, treatment and management. *Br. J. Nurs.*, **2013**, *22*(17), S23-S30. <http://dx.doi.org/10.12968/bjon.2013.22.Sup17.S23> PMID: 24067270
- [9] Narod, S. Can advanced-stage ovarian cancer be cured? *Nat. Rev. Clin. Oncol.*, **2016**, *13*(4), 255-261. <http://dx.doi.org/10.1038/nrclinonc.2015.224> PMID: 26787282
- [10] Orr, B.; Edwards, R.P. Diagnosis and treatment of ovarian cancer. *Hematol. Oncol. Clin. North Am.*, **2018**, *32*(6), 943-964. <http://dx.doi.org/10.1016/j.hoc.2018.07.010> PMID: 30390767
- [11] Eisenhauer, E. Real-world evidence in the treatment of ovarian cancer. *Ann. Oncol.*, **2017**, *28*, viii61-viii65. <http://dx.doi.org/10.1093/annonc/mdx443>
- [12] Mueller, J.J.; Zhou, Q.C.; Iasonos, A.; O'Ceirbhail, R.E.; Alvi, F.A.; El Haraki, A.; Eriksson, A.G.; Gardner, G.J.; Sonoda, Y.; Levine, D.A.; Aghajanian, C.; Chi, D.S.; Abu-Rustum, N.R.; Zivanovic, O. Neoadjuvant chemotherapy and primary debulking surgery utilization for advanced-stage ovarian cancer at a comprehensive cancer center. *Gynecol. Oncol.*, **2016**, *140*(3), 436-442. <http://dx.doi.org/10.1016/j.ygyno.2016.01.008> PMID: 26777991
- [13] Melamed, A.; Hinchcliff, E.M.; Clemmer, J.T.; Bregar, A.J.; Uppal, S.; Bostock, I.; Schorge, J.O.; Del Carmen, M.G.; Rauh-Hain, J.A. Trends in the use of neoadjuvant chemotherapy for advanced ovarian cancer in the United States. *Gynecol. Oncol.*, **2016**, *143*(2), 236-240. <http://dx.doi.org/10.1016/j.ygyno.2016.09.002> PMID: 27612977
- [14] Iorio, G.C.; Martini, S.; Arcadipane, F.; Ricardi, U.; Franco, P. The role of radiotherapy in epithelial ovarian cancer: A literature overview. *Med. Oncol.*, **2019**, *36*(7), 64. <http://dx.doi.org/10.1007/s12032-019-1287-8> PMID: 31165334
- [15] Grunewald, T.; Ledermann, J.A. Targeted therapies for ovarian cancer. *Best Pract. Res. Clin. Obstet. Gynaecol.*, **2017**, *41*, 139-152. <http://dx.doi.org/10.1016/j.bpobgyn.2016.12.001> PMID: 28111228
- [16] Odunsi, K. Immunotherapy in ovarian cancer. *Ann. Oncol.*, **2017**, *28*, viii1-viii7. <http://dx.doi.org/10.1093/annonc/mdx444>
- [17] Mittica, G.; Ghisoni, E.; Giannone, G.; Genta, S.; Aglietta, M.; Sapino, A.; Valabrega, G. PARP inhibitors in ovarian cancer. *Recent Patents Anticancer Drug Discov.*, **2018**, *13*(4), 392-410. <http://dx.doi.org/10.2174/1574892813666180305165256> PMID: 29512470
- [18] Mabuchi, S.; Kuroda, H.; Takahashi, R.; Sasano, T. The PI3K/AKT/mTOR pathway as a therapeutic target in ovarian cancer. *Gynecol. Oncol.*, **2015**, *137*(1), 173-179. <http://dx.doi.org/10.1016/j.ygyno.2015.02.003> PMID: 25677064
- [19] Arend, R.C.; Jackson-Fisher, A.; Jacobs, I.A.; Chou, J.; Monk, B.J. Ovarian cancer: New strategies and emerging targets for the treatment of patients with advanced disease. *Cancer Biol. Ther.*, **2021**, *22*(2), 89-105. <http://dx.doi.org/10.1080/15384047.2020.1868937> PMID: 33427569
- [20] Allemani, C.; Weir, H.K.; Carreira, H.; Harewood, R.; Spika, D.; Wang, X-S. Global surveillance of cancer survival 1995-2009:

- Analysis of individual data for 25 676 887 patients from 279 population-based registries in 67 countries (CONCORD-2). *Lancet*, **2015**, 385, 977-1010.
[http://dx.doi.org/10.1016/S0140-6736\(14\)62038-9](http://dx.doi.org/10.1016/S0140-6736(14)62038-9) PMID: 25467588
- [21] Du, B.; Meenu, M.; Liu, H.; Xu, B. A Concise Review on the Molecular structure and function relationship of β -glucan. *Int. J. Mol. Sci.*, **2019**, 20(16), 4032.
<http://dx.doi.org/10.3390/ijms20164032> PMID: 31426608
- [22] Chan, G.C-F.; Chan, W.K.; Sze, D.M-Y. The effects of β -glucan on human immune and cancer cells. *J. Hematol. Oncol.*, **2009**, 2, 25.
<http://dx.doi.org/10.1186/1756-8722-2-25> PMID: 19515245
- [23] Choromanska, A.; Kulbacka, J.; Harasym, J.; Oledzki, R.; Szewczyk, A.; Saczko, J. High- and low-molecular weight oat beta-glucan reveals antitumor activity in human epithelial lung cancer. *Pathol. Oncol. Res.*, **2018**, 24(3), 583-592.
<http://dx.doi.org/10.1007/s12253-017-0278-3> PMID: 28756506
- [24] Wang, Y.; Harding, S.V.; Thandapilly, S.J.; Tosh, S.M.; Jones, P.J.H.; Ames, N.P. Barley β -glucan reduces blood cholesterol levels via interrupting bile acid metabolism. *Br. J. Nutr.*, **2017**, 118(10), 822-829.
<http://dx.doi.org/10.1017/S0007114517002835> PMID: 29115200
- [25] Geller, A.; Shrestha, R.; Yan, J. Yeast-Derived β -Glucan in Cancer: Novel uses of a traditional therapeutic. *Int. J. Mol. Sci.*, **2019**, 20(15), 3618.
<http://dx.doi.org/10.3390/ijms20153618> PMID: 31344853
- [26] Zhang, Y.; Zhang, M.; Jiang, Y.; Li, X.; He, Y.; Zeng, P.; Guo, Z.; Chang, Y.; Luo, H.; Liu, Y.; Hao, C.; Wang, H.; Zhang, G.; Zhang, L. Lentinan as an immunotherapeutic for treating lung cancer: A review of 12 years clinical studies in China. *J. Cancer Res. Clin. Oncol.*, **2018**, 144(11), 2177-2186.
<http://dx.doi.org/10.1007/s00432-018-2718-1> PMID: 30043277
- [27] McIntosh, M.; Stone, B.A.; Stanisich, V.A. Curdlan and other bacterial (1 \rightarrow 3)-beta-D-glucans. *Appl. Microbiol. Biotechnol.*, **2005**, 68(2), 163-173.
<http://dx.doi.org/10.1007/s00253-005-1959-5> PMID: 15818477
- [28] Vetvicka, V.; Vannucci, L.; Sima, P.; Richter, J. Beta Glucan: Supplement or drug? from laboratory to clinical trials. *Molecules*, **2019**, 24(7), 1251.
<http://dx.doi.org/10.3390/molecules24071251> PMID: 30935016
- [29] Goodridge, H.S.; Wolf, A.J.; Underhill, D.M. Beta-glucan recognition by the innate immune system. *Immunol. Rev.*, **2009**, 230(1), 38-50.
<http://dx.doi.org/10.1111/j.1600-065X.2009.00793.x> PMID: 19594628
- [30] Lukácsi, S.; Nagy-Baló, Z.; Erdei, A.; Sándor, N.; Bajtay, Z. The role of CR3 (CD11b/CD18) and CR4 (CD11c/CD18) in complement-mediated phagocytosis and podosome formation by human phagocytes. *Immunol. Lett.*, **2017**, 189, 64-72.
<http://dx.doi.org/10.1016/j.imlet.2017.05.014> PMID: 28554712
- [31] Xia, Y.; Vetvicka, V.; Yan, J.; Hanikýrová, M.; Mayadas, T.; Ross, G.D. The beta-glucan-binding lectin site of mouse CR3 (CD11b/CD18) and its function in generating a primed state of the receptor that mediates cytotoxic activation in response to iC3b-opsonized target cells. *J. Immunol.*, **1999**, 162(4), 2281-2290.
 PMID: 9973505
- [32] Xue, W.; Kindzelskii, A.L.; Todd, R.F., III; Petty, H.R. Physical association of complement receptor type 3 and urokinase-type plasminogen activator receptor in neutrophil membranes. *J. Immunol.*, **1994**, 152(9), 4630-4640.
 PMID: 8157977
- [33] Chan, G.C.; Chan, W.K.; Sze, D.M. The effects of beta-glucan on human immune and cancer cells. *J. Hematol. Oncol.*, **2009**, 2, 25.
<http://dx.doi.org/10.1186/1756-8722-2-25> PMID: 19515245
- [34] Sahasrabudhe, N.M.; Schols, H.A.; Faas, M.M.; de Vos, P. Arabinoxylan activates Dectin-1 and modulates particulate β -glucan-induced Dectin-1 activation. *Mol. Nutr. Food Res.*, **2016**, 60(2), 458-467.
<http://dx.doi.org/10.1002/mnfr.201500582> PMID: 26394716
- [35] Ariizumi, K.; Shen, G.L.; Shikano, S.; Xu, S.; Ritter, R., III; Kumamoto, T.; Edelbaum, D.; Morita, A.; Bergstresser, P.R.; Takashima, A. Identification of a novel, dendritic cell-associated molecule, dectin-1, by subtractive cDNA cloning. *J. Biol. Chem.*, **2000**, 275(26), 20157-20167.
<http://dx.doi.org/10.1074/jbc.M909512199> PMID: 10779524
- [36] Barton, C.; Vigor, K.; Scott, R.; Jones, P.; Lentfer, H.; Bax, H.J.; Josephs, D.H.; Karagiannis, S.N.; Spicer, J.F. Beta-glucan contamination of pharmaceutical products: How much should we accept? *Cancer Immunol. Immunother.*, **2016**, 65(11), 1289-1301.
<http://dx.doi.org/10.1007/s00262-016-1875-9> PMID: 27473075
- [37] Willment, J.A.; Marshall, A.S.; Reid, D.M.; Williams, D.L.; Wong, S.Y.; Gordon, S.; Brown, G.D. The human beta-glucan receptor is widely expressed and functionally equivalent to murine Dectin-1 on primary cells. *Eur. J. Immunol.*, **2005**, 35(5), 1539-1547.
<http://dx.doi.org/10.1002/eji.200425725> PMID: 15816015
- [38] Brown, G.D. Dectin-1: a signalling non-TLR pattern-recognition receptor. *Nat. Rev. Immunol.*, **2006**, 6(1), 33-43.
<http://dx.doi.org/10.1038/nri1745> PMID: 16341139
- [39] Saijo, S.; Iwakura, Y. Dectin-1 and Dectin-2 in innate immunity against fungi. *Int. Immunol.*, **2011**, 23(8), 467-472.
<http://dx.doi.org/10.1093/intimm/dxr046> PMID: 21677049
- [40] Wakshull, E.; Brunke-Reese, D.; Lindermuth, J.; Fisette, L.; Nathans, R.S.; Crowley, J.J.; Tufts, J.C.; Zimmerman, J.; Mackin, W.; Adams, D.S. PGG-glucan, a soluble beta-(1,3)-glucan, enhances the oxidative burst response, microbicidal activity, and activates an NF-kappa B-like factor in human PMN: evidence for a glycosphingolipid beta-(1,3)-glucan receptor. *Immunopharmacology*, **1999**, 41(2), 89-107.
[http://dx.doi.org/10.1016/S0162-3109\(98\)00059-9](http://dx.doi.org/10.1016/S0162-3109(98)00059-9) PMID: 10102791
- [41] Won, J.S.; Singh, A.K.; Singh, I. Lactosylceramide: a lipid second messenger in neuroinflammatory disease. *J. Neurochem.*, **2007**, 103(Suppl. 1), 180-191.
<http://dx.doi.org/10.1111/j.1471-4159.2007.04822.x> PMID: 17986153
- [42] Vera, J.; Fenutría, R.; Cañadas, O.; Figueras, M.; Mota, R.; Sarrias, M.R.; Williams, D.L.; Casals, C.; Yelamos, J.; Lozano, F. The CD5 ectodomain interacts with conserved fungal cell wall components and protects from zymosan-induced septic shock-like syndrome. *Proc. Natl. Acad. Sci. USA*, **2009**, 106(5), 1506-1511.
<http://dx.doi.org/10.1073/pnas.0805846106> PMID: 19141631
- [43] Sen, G.; Bikah, G.; Venkataraman, C.; Bondada, S. Negative regulation of antigen receptor-mediated signaling by constitutive association of CD5 with the SHP-1 protein tyrosine phosphatase in B-1 B cells. *Eur. J. Immunol.*, **1999**, 29(10), 3319-3328.
[http://dx.doi.org/10.1002/\(SICI\)1521-4141\(199910\)29:10<3319::AID-IMMU3319>3.0.CO;2-9](http://dx.doi.org/10.1002/(SICI)1521-4141(199910)29:10<3319::AID-IMMU3319>3.0.CO;2-9) PMID: 10540344
- [44] Walsh, T.; Casadei, S.; Lee, M.K.; Pennil, C.C.; Nord, A.S.; Thornton, A.M.; Roeb, W.; Agnew, K.J.; Stray, S.M.; Wickramanayake, A.; Norquist, B.; Pennington, K.P.; Garcia, R.L.; King, M.C.; Swisher, E.M. Mutations in 12 genes for inherited ovarian, fallopian tube, and peritoneal carcinoma identified by massively parallel sequencing. *Proc. Natl. Acad. Sci. USA*, **2011**, 108(44), 18032-18037.
<http://dx.doi.org/10.1073/pnas.1115052108> PMID: 22006311
- [45] Moschetta, M.; George, A.; Kaye, SB; Banerjee, S BRCA somatic mutations and epigenetic BRCA modifications in serous ovarian cancer. *Annal Oncol* : **2016**, 27, 1449-1455.

- <http://dx.doi.org/10.1093/annonc/mdw142>
- [46] Toss, A.; Tomasello, C.; Razzaboni, E.; Contu, G.; Grandi, G.; Cagnacci, A.; Schilder, R.J.; Cortesi, L. Hereditary ovarian cancer: not only BRCA 1 and 2 genes. *BioMed Res. Int.*, **2015**, *2015*, 341723.
<http://dx.doi.org/10.1155/2015/341723> PMID: 26075229
- [47] Sadoughi, F.; Hallajzadeh, J.; Asemi, Z.; Mansournia, M.A.; Alemi, F.; Yousefi, B. Signaling pathways involved in cell cycle arrest during the DNA breaks. *DNA Repair (Amst.)*, **2021**, *98*, 103047.
<http://dx.doi.org/10.1016/j.dnarep.2021.103047> PMID: 33454524
- [48] Sadoughi, F.; Mirsafaei, L.; Dana, P.M.; Hallajzadeh, J.; Asemi, Z.; Mansournia, M.A.; Montazer, M.; Hosseinpour, M.; Yousefi, B. The role of DNA damage response in chemo- and radio-resistance of cancer cells: Can DDR inhibitors solve the problem? *DNA Repair (Amst.)*, **2021**, *101*, 103074.
<http://dx.doi.org/10.1016/j.dnarep.2021.103074> PMID: 33640757
- [49] Chatterjee, N.; Walker, G.C. Mechanisms of DNA damage, repair, and mutagenesis. *Environ. Mol. Mutagen.*, **2017**, *58*(5), 235-263.
<http://dx.doi.org/10.1002/em.22087> PMID: 28485537
- [50] Jackson, S.P.; Bartek, J. The DNA-damage response in human biology and disease. *Nature*, **2009**, *461*(7267), 1071-1078.
<http://dx.doi.org/10.1038/nature08467> PMID: 19847258
- [51] Coussens, L.M.; Werb, Z. Inflammation and cancer. *Nature*, **2002**, *420*(6917), 860-867.
<http://dx.doi.org/10.1038/nature01322> PMID: 12490959
- [52] Kuper, H.; Adami, H.O.; Trichopoulos, D. Infections as a major preventable cause of human cancer. *J. Intern. Med.*, **2000**, *248*(3), 171-183.
<http://dx.doi.org/10.1046/j.1365-2796.2000.00742.x> PMID: 10971784
- [53] Dhillon, A.S.; Hagan, S.; Rath, O.; Kolch, W. MAP kinase signalling pathways in cancer. *Oncogene*, **2007**, *26*(22), 3279-3290.
<http://dx.doi.org/10.1038/sj.onc.1210421> PMID: 17496922
- [54] Schulze-Osthoff, K.; Ferrari, D.; Riehemann, K.; Wesselborg, S. Regulation of NF- κ B activation by MAP kinase cascades. *Immunobiology*, **1997**, *198*(1-3), 35-49.
[http://dx.doi.org/10.1016/S0171-2985\(97\)80025-3](http://dx.doi.org/10.1016/S0171-2985(97)80025-3) PMID: 9442376
- [55] Chou, C.H.; Wei, L.H.; Kuo, M.L.; Huang, Y.J.; Lai, K.P.; Chen, C.A.; Hsieh, C.Y. Up-regulation of interleukin-6 in human ovarian cancer cell via a Gi/PI3K-Akt/NF-kappaB pathway by lysophosphatidic acid, an ovarian cancer-activating factor. *Carcinogenesis*, **2005**, *26*(1), 45-52.
<http://dx.doi.org/10.1093/carcin/bgh301> PMID: 15471896
- [56] Grivennikov, S.I.; Greten, F.R.; Karin, M. Immunity, inflammation, and cancer. *Cell*, **2010**, *140*(6), 883-899.
<http://dx.doi.org/10.1016/j.cell.2010.01.025> PMID: 20303878
- [57] Savant, S.S.; Sriramkumar, S.; O'Hagan, H.M. The role of inflammation and inflammatory mediators in the development, progression, metastasis, and chemoresistance of epithelial ovarian cancer. *Cancers (Basel)*, **2018**, *10*(8), 251.
<http://dx.doi.org/10.3390/cancers10080251> PMID: 30061485
- [58] Kroeger, P.T., Jr; Drapkin, R. Pathogenesis and heterogeneity of ovarian cancer. *Curr. Opin. Obstet. Gynecol.*, **2017**, *29*(1), 26-34.
<http://dx.doi.org/10.1097/GCO.0000000000000340> PMID: 27898521
- [59] Petrovská, M.; Dimitrov, D.G.; Michael, S.D. Quantitative changes in macrophage distribution in normal mouse ovary over the course of the estrous cycle examined with an image analysis system. *Am. J. Reprod. Immunol.*, **1996**, *36*(3), 175-183.
<http://dx.doi.org/10.1111/j.1600-0897.1996.tb00159.x> PMID: 8874714
- [60] Gupta, V.; Yull, F.; Khabele, D. Bipolar tumor-associated macrophages in ovarian cancer as targets for therapy. *Cancers (Basel)*, **2018**, *10*(10), 366.
<http://dx.doi.org/10.3390/cancers10100366> PMID: 30274280
- [61] Chumduri, C.; Gurumurthy, R.K.; Zadora, P.K.; Mi, Y.; Meyer, T.F. Chlamydia infection promotes host DNA damage and proliferation but impairs the DNA damage response. *Cell Host Microbe*, **2013**, *13*(6), 746-758.
<http://dx.doi.org/10.1016/j.chom.2013.05.010> PMID: 23768498
- [62] Ingerslev, K.; Hogdall, E.; Schnack, T.H.; Skovrider-Ruminski, W.; Hogdall, C.; Blaakaer, J. The potential role of infectious agents and pelvic inflammatory disease in ovarian carcinogenesis. *Infect. Agent. Cancer*, **2017**, *12*, 25.
<http://dx.doi.org/10.1186/s13027-017-0134-9> PMID: 28529540
- [63] Gunderson, C.C.; Ding, K.; Dvorak, J.; Moore, K.N.; McMeekin, D.S.; Benbrook, D.M. The pro-inflammatory effect of obesity on high grade serous ovarian cancer. *Gynecol. Oncol.*, **2016**, *143*(1), 40-45.
<http://dx.doi.org/10.1016/j.ygyno.2016.07.103> PMID: 27423378
- [64] Sayasneh, A.; Tsivos, D.; Crawford, R. Endometriosis and ovarian cancer: a systematic review. *ISRN Obstet. Gynecol.*, **2011**, *2011*, 140310.
<http://dx.doi.org/10.5402/2011/140310> PMID: 21789283
- [65] Harris, H.R.; Terry, K.L. Polycystic ovary syndrome and risk of endometrial, ovarian, and breast cancer: a systematic review. *Fertil. Res. Pract.*, **2016**, *2*, 14.
<http://dx.doi.org/10.1186/s40738-016-0029-2> PMID: 28620541
- [66] Pistritto, G.; Trisciuglio, D.; Ceci, C.; Garufi, A.; D'Orazi, G. Apoptosis as anticancer mechanism: function and dysfunction of its modulators and targeted therapeutic strategies. *Aging (Albany NY)*, **2016**, *8*(4), 603-619.
<http://dx.doi.org/10.18632/aging.100934> PMID: 27019364
- [67] Elmore, S. Apoptosis: a review of programmed cell death. *Toxicol. Pathol.*, **2007**, *35*(4), 495-516.
<http://dx.doi.org/10.1080/01926230701320337> PMID: 17562483
- [68] D'Arcy, M.S. Cell death: A review of the major forms of apoptosis, necrosis and autophagy. *Cell Biol. Int.*, **2019**, *43*(6), 582-592.
<http://dx.doi.org/10.1002/cbin.11137> PMID: 30958602
- [69] Avontuur, J.A.; Tutein Nolthenius, R.P.; Buijk, S.L.; Kanhai, K.J.; Bruining, H.A. Effect of L-NAME, an inhibitor of nitric oxide synthesis, on cardiopulmonary function in human septic shock. *Chest*, **1998**, *113*(6), 1640-1646.
<http://dx.doi.org/10.1378/chest.113.6.1640> PMID: 9631805
- [70] Browning, L.; Patel, M.R.; Horvath, E.B.; Tawara, K.; Jorczyk, C.L. IL-6 and ovarian cancer: inflammatory cytokines in promotion of metastasis. *Cancer Manag. Res.*, **2018**, *10*, 6685-6693.
<http://dx.doi.org/10.2147/CMAR.S179189> PMID: 30584363
- [71] Malone, J.M.; Saed, G.M.; Diamond, M.P.; Sokol, R.J.; Munkarah, A.R. The effects of the inhibition of inducible nitric oxide synthase on angiogenesis of epithelial ovarian cancer. *Am. J. Obstet. Gynecol.*, **2006**, *194*(4), 1110-1116.
<http://dx.doi.org/10.1016/j.ajog.2005.12.019> PMID: 16580304
- [72] Role of the PI3K/AKT/mTOR signaling pathway in ovarian cancer: Biological and therapeutic significance. Ediriweera, M.K.; Tennekoon, K.H.; Samarakoon, S.R., Eds.; *Seminars in cancer biology*; Elsevier, **2019**.
- [73] Meng, Q.; Xia, C.; Fang, J.; Rojanasakul, Y.; Jiang, B-H. Role of PI3K and AKT specific isoforms in ovarian cancer cell migration, invasion and proliferation through the p70S6K1 pathway. *Cell. Signal.*, **2006**, *18*(12), 2262-2271.
<http://dx.doi.org/10.1016/j.cellsig.2006.05.019> PMID: 16839745
- [74] Montero, J.C.; Chen, X.; Ocaña, A.; Pandiella, A. Predominance of mTORC1 over mTORC2 in the regulation of proliferation of ovarian cancer cells: therapeutic implications. *Mol. Cancer Ther.*, **2012**, *11*(6), 1342-1352.
<http://dx.doi.org/10.1158/1535-7163.MCT-11-0723> PMID: 22496482

- [75] Li, S.; Lv, M.; Qiu, S.; Meng, J.; Liu, W.; Zuo, J.; Yang, L. NF- κ B p65 promotes ovarian cancer cell proliferation and migration via regulating mortalin. *J. Cell. Mol. Med.*, **2019**, *23*(6), 4338-4348. <http://dx.doi.org/10.1111/jcmm.14325> PMID: 30983127
- [76] Xiao, X.; Yang, G.; Bai, P.; Gui, S.; Nyuyen, T.M.; Mercado-Uribe, I.; Yang, M.; Zou, J.; Li, Q.; Xiao, J.; Chang, B.; Liu, G.; Wang, H.; Liu, J. Inhibition of nuclear factor-kappa B enhances the tumor growth of ovarian cancer cell line derived from a low-grade papillary serous carcinoma in p53-independent pathway. *BMC Cancer*, **2016**, *16*, 582. <http://dx.doi.org/10.1186/s12885-016-2617-2> PMID: 27484466
- [77] Yang, G.; Xiao, X.; Rosen, D.G.; Cheng, X.; Wu, X.; Chang, B. The biphasic role of NF-kappaB in progression and chemoresistance of ovarian cancer. *Clin Cancer Res : an official journal of the American Association for Cancer Research*, **2011**, *17*, 2181-2194.
- [78] Guan, X.; Chen, S.; Liu, Y.; Wang, L.L.; Zhao, Y.; Zong, Z.H. PUM1 promotes ovarian cancer proliferation, migration and invasion. *Biochem. Biophys. Res. Commun.*, **2018**, *497*(1), 313-318. <http://dx.doi.org/10.1016/j.bbrc.2018.02.078> PMID: 29428722
- [79] Hou, X.S.; Han, C.Q.; Zhang, W. MiR-1182 inhibited metastasis and proliferation of ovarian cancer by targeting hTERT. *Eur. Rev. Med. Pharmacol. Sci.*, **2018**, *22*(6), 1622-1628. PMID: 29630105
- [80] Jiang, J.H.; Lv, Q.Y.; Yi, Y.X.; Liao, J.; Wang, X.W.; Zhang, W. MicroRNA-200a promotes proliferation and invasion of ovarian cancer cells by targeting PTEN. *Eur. Rev. Med. Pharmacol. Sci.*, **2018**, *22*(19), 6260-6267. PMID: 30338796
- [81] Wang, L.; Yan, W.; Li, X.; Liu, Z.; Tian, T.; Chen, T.; Zou, L.; Cui, Z. S100A10 silencing suppresses proliferation, migration and invasion of ovarian cancer cells and enhances sensitivity to carboplatin. *J. Ovarian Res.*, **2019**, *12*(1), 113. <http://dx.doi.org/10.1186/s13048-019-0592-3> PMID: 31739800
- [82] Xiang, G.; Cheng, Y. MiR-126-3p inhibits ovarian cancer proliferation and invasion via targeting PLXNB2. *Reprod. Biol.*, **2018**, *18*(3), 218-224. <http://dx.doi.org/10.1016/j.repbio.2018.07.005> PMID: 30054097
- [83] Yang, B.; Sun, L.; Liang, L. MiRNA-802 suppresses proliferation and migration of epithelial ovarian cancer cells by targeting YWHAZ. *J. Ovarian Res.*, **2019**, *12*(1), 100. <http://dx.doi.org/10.1186/s13048-019-0576-3> PMID: 31640760
- [84] Zhan, F.L.; Chen, C.F.; Yao, M.Z. LncRNA TUG1 facilitates proliferation, invasion and stemness of ovarian cancer cell via miR-186-5p/ZEB1 axis. *Cell Biochem. Funct.*, **2020**, *38*(8), 1069-1078. <http://dx.doi.org/10.1002/cbf.3544> PMID: 32390141
- [85] Zheng, F.; Xiao, X.; Wang, C. the effect of pth1 on ovarian cancer cell proliferation and apoptosis. *Cancer Biother. Radiopharm.*, **2019**, *34*(2), 103-109. <http://dx.doi.org/10.1089/cbr.2018.2626> PMID: 30523702
- [86] Zheng, Z.J.; Liu, Y.; Wang, H.J.; Pang, W.W.; Wang, Y. LncRNA SNHG17 promotes proliferation and invasion of ovarian cancer cells through up-regulating FOXA1. *Eur. Rev. Med. Pharmacol. Sci.*, **2020**, *24*(18), 9282-9289. PMID: 33015769
- [87] Vetvicka, V.; Teplyakova, T.V.; Shintyapina, A.B.; Korolenko, T.A. effects of medicinal fungi-derived β -glucan on tumor progression. *J. Fungi (Basel)*, **2021**, *7*(4), 250. <http://dx.doi.org/10.3390/jof7040250> PMID: 33806255
- [88] Fang, J.; Wang, Y.; Lv, X.; Shen, X.; Ni, X.; Ding, K. Structure of a β -glucan from *Grifola frondosa* and its antitumor effect by activating Dectin-1/Syk/NF- κ B signaling. *Glycoconj. J.*, **2012**, *29*(5-6), 365-377. <http://dx.doi.org/10.1007/s10719-012-9416-z> PMID: 22744837
- [89] Chaichian, S.; Moazzami, B.; Sadoughi, F.; Haddad Kashani, H.; Zaroudi, M.; Asemi, Z. Functional activities of beta-glucans in the prevention or treatment of cervical cancer. *J. Ovarian Res.*, **2020**, *13*(1), 24. <http://dx.doi.org/10.1186/s13048-020-00626-7> PMID: 32138756
- [90] Legentil, L.; Paris, F.; Ballet, C.; Trouvelot, S.; Daire, X.; Vetvicka, V.; Ferrières, V. Molecular interactions of β -(1 \rightarrow 3)-glucans with their receptors. *Molecules*, **2015**, *20*(6), 9745-9766. <http://dx.doi.org/10.3390/molecules20069745> PMID: 26023937
- [91] Baldassano, S.; Accardi, G.; Vasto, S. Beta-glucans and cancer: The influence of inflammation and gut peptide. *Eur. J. Med. Chem.*, **2017**, *142*, 486-492. <http://dx.doi.org/10.1016/j.ejmech.2017.09.013> PMID: 28964548
- [92] Olson, E.J.; Standing, J.E.; Griego-Harper, N.; Hoffman, O.A.; Limper, A.H. Fungal beta-glucan interacts with vitronectin and stimulates tumor necrosis factor alpha release from macrophages. *Infect. Immun.*, **1996**, *64*(9), 3548-3554. <http://dx.doi.org/10.1128/iai.64.9.3548-3554.1996> PMID: 8751898
- [93] Brown, G.D.; Herre, J.; Williams, D.L.; Willment, J.A.; Marshall, A.S.; Gordon, S. Dectin-1 mediates the biological effects of β -glucans. *J. Exp. Med.*, **2003**, *197*(9), 1119-1124. <http://dx.doi.org/10.1084/jem.20021890> PMID: 12719478
- [94] Engstad, C.S.; Engstad, R.E.; Olsen, J-O.; Østerud, B. The effect of soluble β -1,3-glucan and lipopolysaccharide on cytokine production and coagulation activation in whole blood. *Int. Immunopharmacol.*, **2002**, *2*(11), 1585-1597. [http://dx.doi.org/10.1016/S1567-5769\(02\)00134-0](http://dx.doi.org/10.1016/S1567-5769(02)00134-0) PMID: 12433059
- [95] Estrada, A.; Yun, C-H.; Van Kessel, A.; Li, B.; Hauta, S.; Laarveld, B. Immunomodulatory activities of oat β -glucan *in vitro* and *in vivo*. *Microbiol. Immunol.*, **1997**, *41*(12), 991-998. <http://dx.doi.org/10.1111/j.1348-0421.1997.tb01959.x> PMID: 9492185
- [96] Hahn, P.Y.; Evans, S.E.; Kottom, T.J.; Standing, J.E.; Pagano, R.E.; Limper, A.H. Pneumocystis carinii cell wall β -glucan induces release of macrophage inflammatory protein-2 from alveolar epithelial cells via a lactosylceramide-mediated mechanism. *J. Biol. Chem.*, **2003**, *278*(3), 2043-2050. <http://dx.doi.org/10.1074/jbc.M209715200> PMID: 12419803
- [97] Lin, Y-L.; Lee, S-S.; Hou, S-M.; Chiang, B-L. Polysaccharide purified from *Ganoderma lucidum* induces gene expression changes in human dendritic cells and promotes T helper 1 immune response in BALB/c mice. *Mol. Pharmacol.*, **2006**, *70*(2), 637-644. <http://dx.doi.org/10.1124/mol.106.022327> PMID: 16670374
- [98] Steimbach, L.; Borgmann, A.V.; Gomar, G.G.; Hoffmann, L.V.; Rutckeviski, R.; de Andrade, D.P. *Fungal beta-glucans as adjuvants for treating cancer patients-A Systematic Review of Clinical Trials*; Clin Nut, **2020**.
- [99] Suram, S.; Brown, G.D.; Ghosh, M.; Gordon, S.; Loper, R.; Taylor, P.R.; Akira, S.; Uematsu, S.; Williams, D.L.; Leslie, C.C. Regulation of cytosolic phospholipase A2 activation and cyclooxygenase 2 expression in macrophages by the β -glucan receptor. *J. Biol. Chem.*, **2006**, *281*(9), 5506-5514. <http://dx.doi.org/10.1074/jbc.M509824200> PMID: 16407295
- [100] Akramiene, D.; Kondrotas, A.; Didziapetriene, J.; Kevelaitis, E. Effects of beta-glucans on the immune system. *Medicina (Kaunas)*, **2007**, *43*(8), 597-606. <http://dx.doi.org/10.3390/medicina43080076> PMID: 17895634
- [101] de Graaff, P.; Govers, C.; Wichers, H.J.; Debets, R. Consumption of β -glucans to spice up T cell treatment of tumors: a review. *Expert Opin. Biol. Ther.*, **2018**, *18*(10), 1023-1040. <http://dx.doi.org/10.1080/14712598.2018.1523392> PMID: 30221551
- [102] Goyal, S.; Castrillón-Betancur, J.C.; Klaile, E.; Slevogt, H. The interaction of human pathogenic fungi With C-Type Lectin Receptors. *Front. Immunol.*, **2018**, *9*, 1261.

- <http://dx.doi.org/10.3389/fimmu.2018.01261> PMID: 29915598
- [103] Sun, W.-K.; Lu, X.; Li, X.; Sun, Q.-Y.; Su, X.; Song, Y.; Sun, H.M.; Shi, Y. Dectin-1 is inducible and plays a crucial role in Aspergillus-induced innate immune responses in human bronchial epithelial cells. *Eur. J. Clin. Microbiol. Infect. Dis.*, **2012**, *31*(10), 2755-2764. <http://dx.doi.org/10.1007/s10096-012-1624-8> PMID: 22562430
- [104] Liang, J.; Melican, D.; Cafro, L.; Palace, G.; Fiset, L.; Armstrong, R.; Patchen, M.L. Enhanced clearance of a multiple antibiotic resistant *Staphylococcus aureus* in rats treated with PGG-glucan is associated with increased leukocyte counts and increased neutrophil oxidative burst activity. *Int. J. Immunopharmacol.*, **1998**, *20*(11), 595-614. [http://dx.doi.org/10.1016/S0192-0561\(98\)00007-1](http://dx.doi.org/10.1016/S0192-0561(98)00007-1) PMID: 9848393
- [105] Bose, N.; Wurst, L.R.; Chan, A.S.; Dudney, C.M.; LeRoux, M.L.; Danielson, M.E.; Will, P.M.; Nodland, S.E.; Patchen, M.L.; Dalle Lucca, J.J.; Lebeda, F.J.; Vasilakos, J.P. Differential regulation of oxidative burst by distinct β -glucan-binding receptors and signaling pathways in human peripheral blood mononuclear cells. *Glycobiology*, **2014**, *24*(4), 379-391. <http://dx.doi.org/10.1093/glycob/cwu005> PMID: 24440830
- [106] Tian, J.; Ma, J.; Ma, K.; Guo, H.; Baidoo, S.E.; Zhang, Y.; Yan, J.; Lu, L.; Xu, H.; Wang, S. β -Glucan enhances antitumor immune responses by regulating differentiation and function of monocytic myeloid-derived suppressor cells. *Eur. J. Immunol.*, **2013**, *43*(5), 1220-1230. <http://dx.doi.org/10.1002/eji.201242841> PMID: 23424024
- [107] Vlahopoulos, S.A. Aberrant control of NF- κ B in cancer permits transcriptional and phenotypic plasticity, to curtail dependence on host tissue: molecular mode. *Cancer Biol. Med.*, **2017**, *14*(3), 254-270. <http://dx.doi.org/10.20892/j.issn.2095-3941.2017.0029> PMID: 28884042
- [108] Harbort, C.J.; Soeiro-Pereira, P.V.; von Bernuth, H.; Kaindl, A.M.; Costa-Carvalho, B.T.; Condino-Neto, A.; Reichenbach, J.; Roesler, J.; Zychlinsky, A.; Amulic, B. Neutrophil oxidative burst activates ATM to regulate cytokine production and apoptosis. *Blood*, **2015**, *126*(26), 2842-2851. <http://dx.doi.org/10.1182/blood-2015-05-645424> PMID: 26491069
- [109] Wang, N.; Liu, H.; Liu, G.; Li, M.; He, X.; Yin, C.; Tu, Q.; Shen, X.; Bai, W.; Wang, Q.; Tao, Y.; Yin, H. Yeast β -D-glucan exerts antitumor activity in liver cancer through impairing autophagy and lysosomal function, promoting reactive oxygen species production and apoptosis. *Redox Biol.*, **2020**, *32*, 101495. <http://dx.doi.org/10.1016/j.redox.2020.101495> PMID: 32171725
- [110] Kim, M.J.; Hong, S.Y.; Kim, S.K.; Cheong, C.; Park, H.J.; Chun, H.K. β -Glucan enhanced apoptosis in human colon cancer cells SNU-C4. *Nut Res practice.*, **2009**, *3*, 180-184.
- [111] Gu, Y.Y.; Chen, M.H.; May, B.H.; Liao, X.Z.; Liu, J.H.; Tao, L.T. Matrine induces apoptosis in multiple colorectal cancer cell lines *in vitro* and inhibits tumour growth with minimum side effects *in vivo* via Bcl-2 and caspase-3. *Phytomedicine: International journal of phytotherapy and phytopharmacology*, **2018**, *51*, 214-225. <http://dx.doi.org/10.1016/j.phymed.2018.10.004>
- [112] Saravanakumar, K.; Jeevithan, E.; Hu, X.; Chelliah, R.; Oh, D.H.; Wang, M.H. Enhanced anti-lung carcinoma and anti-biofilm activity of fungal molecules mediated biogenic zinc oxide nanoparticles conjugated with β -D-glucan from barley. *J. Photochem. Photobiol. B*, **2020**, *203*, 111728. <http://dx.doi.org/10.1016/j.jphotobiol.2019.111728> PMID: 31864088
- [113] Thomas, M.; Sadjadian, P.; Kollmeier, J.; Lowe, J.; Mattson, P.; Trout, J.R.; Gargano, M.; Patchen, M.L.; Walsh, R.; Beliveau, M.; Marier, J.F.; Bose, N.; Gorden, K.; Schneller, F., III A randomized, open-label, multicenter, phase II study evaluating the efficacy and safety of BTH1677 (1,3-1,6 beta glucan; Imprime PGG) in combination with cetuximab and chemotherapy in patients with advanced non-small cell lung cancer. *Invest. New Drugs*, **2017**, *35*(3), 345-358. <http://dx.doi.org/10.1007/s10637-017-0450-3> PMID: 28303530
- [114] Li, X.J.; Jia, Y.J.; Chen, L. Clinical observation of thermotherapy combined with thoracic injection of lentinan in treatment of cancerous hydrothorax of patients with lung cancer. *Zhongguo Zhong Xi Yi Jie He Za Zhi*, **2011**, *31*(8), 1062-1065. [Clinical observation of thermotherapy combined with thoracic injection of lentinan in treatment of cancerous hydrothorax of patients with lung cancer]. PMID: 21910335
- [115] Ostadrahimi, A.; Ziaei, J.E.; Esfahani, A.; Jafarabadi, M.A.; Movassaghpourakbari, A.; Farrin, N. Effect of beta glucan on white blood cell counts and serum levels of IL-4 and IL-12 in women with breast cancer undergoing chemotherapy: a randomized double-blind placebo-controlled clinical trial. *Asian Pac. J. Cancer Prev.*, **2014**, *15*(14), 5733-5739. <http://dx.doi.org/10.7314/APJCP.2014.15.14.5733> PMID: 25081694
- [116] Kataoka, H.; Shimura, T.; Mizoshita, T.; Kubota, E.; Mori, Y.; Mizushima, T.; Wada, T.; Ogasawara, N.; Tanida, S.; Sasaki, M.; Togawa, S.; Sano, H.; Hirata, Y.; Ikai, M.; Mochizuki, H.; Seno, K.; Itoh, S.; Kawai, T.; Joh, T. Lentinan with S-1 and paclitaxel for gastric cancer chemotherapy improve patient quality of life. *Hepatogastroenterology*, **2009**, *56*(90), 547-550. PMID: 19579640
- [117] Tari, K.; Satake, I.; Nakagomi, K.; Ozawa, K.; Oowada, F.; Higashi, Y.; Negishi, T.; Yamada, T.; Saito, H.; Yoshida, K. Effect of lentinan for advanced prostate carcinoma. *Hinyokika Kyo*, **1994**, *40*(2), 119-123. PMID: 8128920
- [118] Zhang, Y.; Li, S.; Wang, X.; Zhang, L.; Cheung, P.C. Advances in lentinan: isolation, structure, chain conformation and bioactivities. *Food Hydrocoll.*, **2011**, *25*, 196-206. <http://dx.doi.org/10.1016/j.foodhyd.2010.02.001>
- [119] Fujimoto, K.; Tomonaga, M.; Goto, S. A case of recurrent ovarian cancer successfully treated with adoptive immunotherapy and lentinan. *Anticancer Res.*, **2006**, *26*(6A), 4015-4018. PMID: 17195451
- [120] Shimizu, Y.; Hasumi, K.; Chen, J.; Hirai, Y.; Nakayama, K.; Teshima, H. Successful treatment of a patient with recurrent ovarian cancer by lentinan combined with intraarterial 5FU. *Nihon Gan Chiryo Gakkai shi.*, **1989**, *24*, 647-651.
- [121] Guo, L.-Y.; Zhang, S.-Y.; Chen, C.; Zeng, H.-X.; Li, F.-Y.; Xu, Q.-X. Lentinan combined with cisplatin and paclitaxel in the treatment of patients with ovarian cancer with ascites. *Eur. J. Gynaecol. Oncol.*, **2018**, *39*, 615-620.
- [122] Zhang, M.; Zhang, Y.; Zhang, L.; Tian, Q. Mushroom polysaccharide lentinan for treating different types of cancers: A review of 12 years clinical studies in China. *Prog. Mol. Biol. Transl. Sci.*, **2019**, *163*, 297-328. <http://dx.doi.org/10.1016/bs.pmbts.2019.02.013> PMID: 31030752
- [123] Liu, X.-d.; Li, M.; Li, W.-x.; Wang, Q.-y.; Zhang, H.-x. Combined effect of lentinan and cisplatin on cytokines IL-6, TNF- α , and TGF- β in tumor therapy. *Int J Polymer Sci.*, **2019**, *2019*.
- [124] Kony, D.B.; Damm, W.; Stoll, S.; van Gunsteren, W.F.; Hünenberger, P.H. Explicit-solvent molecular dynamics simulations of the polysaccharide schizophyllan in water. *Biophys. J.*, **2007**, *93*(2), 442-455. <http://dx.doi.org/10.1529/biophysj.106.086116> PMID: 17237195
- [125] Chen, J.T.; Hasumi, K.; Masubuchi, K. Interferon-alpha, interferon-gamma and sizofiran in the adjuvant therapy in ovarian cancer—a preliminary trial. *Biotherapy*, **1992**, *5*(4), 275-280.

- <http://dx.doi.org/10.1007/BF02179044> PMID: 1290723
- [126] Hoshino, T.; Suzuki, Y.; Takeichi, M.; Adachi, T.; Takayama, M. Combined effects of sizofiran and rG-CSF on myelosuppression in cancer chemotherapy. *Nippon Sanka Fujinka Gakkai Zasshi*, **1996**, *48*(3), 206-212. PMID: 8721055
- [127] Inoue, M.; Tanaka, Y.; Sugita, N.; Yamasaki, M.; Yamanaka, T.; Minagawa, J.; Nakamuro, K.; Tani, T.; Okudaira, Y.; Karita, T. Improvement of long-term prognosis in patients with ovarian cancers by adjuvant sizofiran immunotherapy: a prospective randomized controlled study. *Biotherapy*, **1993**, *6*(1), 13-18. <http://dx.doi.org/10.1007/BF01877381> PMID: 8507540
- [128] Sugiyama, T.; Nishida, T.; Kumagai, S.; Imaishi, K.; Ushijima, K.; Kataoka, A.; Yakushiji, M. Combination treatment with cisplatin and schizophyllan for 7,12-dimethylbenz(a)anthracene-induced rat ovarian adenocarcinoma. *J Obstet Gynaecol (Tokyo 1995)*, **1995**, *21*(5), 521-527. <http://dx.doi.org/10.1111/j.1447-0756.1995.tb01047.x> PMID: 8542479
- [129] Chen, J-T.; Hasumi, K. Activation of peritoneal macrophages in patients with gynecological malignancies by sizofiran and recombinant interferon- γ . *Biotherapy*, **1993**, *6*(3), 189-194. <http://dx.doi.org/10.1007/BF01878080> PMID: 8292460
- [130] Yallapu, M.M.; Dobberpuhl, M.R.; Maher, D.M.; Jaggi, M.; Chauhan, S.C. Design of curcumin loaded cellulose nanoparticles for prostate cancer. *Curr. Drug Metab.*, **2012**, *13*(1), 120-128. <http://dx.doi.org/10.2174/138920012798356952> PMID: 21892919
- [131] Picaud, L.; Thibault, B.; Mery, E.; Ouali, M.; Martinez, A.; Delord, J-P.; Couderc, B.; Ferron, G. Evaluation of the effects of hyaluronic acid-carboxymethyl cellulose barrier on ovarian tumor progression. *J. Ovarian Res.*, **2014**, *7*, 40. <http://dx.doi.org/10.1186/1757-2215-7-40> PMID: 24739440
- [132] Münster, L.; Fojtů, M.; Capáková, Z.; Vaculovič, T.; Tvrdoňová, M.; Kuřitka, I.; Masařík, M.; Vicha, J. Selectively oxidized cellulose with adjustable molecular weight for controlled release of platinum anticancer drugs. *Biomacromolecules*, **2019**, *20*(4), 1623-1634. <http://dx.doi.org/10.1021/acs.biomac.8b01807> PMID: 30794396
- [133] Kadam, S.U.; Tiwari, B.K.; O'Donnell, C.P. Extraction, structure and biofunctional activities of laminarin from brown algae. *Int. J. Food Sci. Technol.*, **2015**, *50*, 24-31. <http://dx.doi.org/10.1111/ijfs.12692>
- [134] Bae, H.; Song, G.; Lee, J.Y.; Hong, T.; Chang, M.J.; Lim, W. Laminarin-derived from brown algae suppresses the growth of ovarian cancer cells via mitochondrial dysfunction and er stress. *Mar. Drugs*, **2020**, *18*(3), 18. <http://dx.doi.org/10.3390/md18030152> PMID: 32182828
- [135] Gockley, A.; Wright, A. living through ovarian cancer treatment: acute and long-term toxicities of chemotherapy for Advanced-Stage Disease. *Hematol. Oncol. Clin. North Am.*, **2018**, *32*(6), 1073-1085. <http://dx.doi.org/10.1016/j.hoc.2018.07.009> PMID: 30390761
- [136] Cheung, N-K.V.; Modak, S.; Vickers, A.; Knuckles, B. Orally administered β -glucans enhance anti-tumor effects of monoclonal antibodies. *Cancer Immunol. Immunother.*, **2002**, *51*(10), 557-564. <http://dx.doi.org/10.1007/s00262-002-0321-3> PMID: 12384807